Introduction

Over the past few years, cancer has been the root cause of disease mortality in the world. Among different variants of cancer, lung cancer (LC) is the most dangerous and leading category of cancer worldwide (1,2). According to the World Health Organization, approximately 2.09 million people suffered from this disease, and it was caused around 1.76 million of mortality among them in the year 2018 (1). LC originates from the alteration of healthy cells into malignant cells in several biochemical processes that typically grow from pre-cancerous lesions to nonbenign tumors, lumps, and neoplasms. This feature, with the fast formation of abnormal cells, raises outside their typical borders. These cancer cells later may invade adjacent organs of the body from the lungs and spread over other parts of the body. This invasion refers to metastasizing, which promotes fatalities from cancer. LC is predominantly caused by two major clusters of histological patterns (2): oat cell or small cell lung cancer (SCLC) and non-small cell lung cancer (NSCLC). 80–85% of LC are NSCLC, whereas the rest of the 10–15% are oat cell type (2). NSCLC is subcategorized in several types following their histological patterns, such as adenocarcinoma, squamous cell carcinoma, large cell...
carcinoma, adenosquamous carcinoma, and sarcomatoid carcinoma (3,4). The roots of adenocarcinoma and squamous cell carcinoma are typically formed in mucous, whereas large cell carcinoma origins in the squamous cells. However, large cell carcinoma intends to spread more rapidly compared to other types of NSCLC due to its aggressive histological patterns. Adenosquamous carcinoma and sarcomatoid carcinoma are fewer common subspecies of LC. Typically, tobacco smoking is the primary cause (80–90%) of promoting all these patterns of LC and the cause of 80–90% of deaths per year (5). Carcinoid tumors, lymphomas, adenoid cystic carcinomas, sarcomas, or rooted from other organs (e.g., breast, pancreas, kidney, etc.) cause fewer than 5% of malignancy in lung cells (6).

The four following significant intrinsic and extrinsic factors have been identified, causing LC so far by the International Agency for Research on Cancer (IARC) (7): (I) a person's genetic influences, (II) physical carcinogens such as ionizing and ultraviolet (UV) radiation, (III) toxic chemical (e.g., tobacco, a food toxic, asbestos, and drinking water contaminants (e.g., arsenic); and (IV) organic carcinogens, such as infections from specific microorganisms (e.g., viruses, bacteria, or parasites). Apart from these, aging leads to the risk of cancers as the cellular reparation mechanisms get inactive with the increment of oldness. Besides, the ingestion of alcohol and tobacco smoke, unhealthy diet, inactive lifestyle, and cancer-causing chronic infections due to Hepatitis B and C viruses, Human papillomavirus (HPV), Helicobacter pylori, and Epstein-Barr virus are the key danger factors to activate LC (8-11).

LC can be prevented by following a healthy lifestyle such as alcohol and tobacco avoidance, a healthy diet, and physical activities (12). Also, the regular screening of lung can reduce the severity of LC. For instance, the results of CT and X-ray in an earlier screening test for the stage I diagnosis of LC was positive for the 63% and 47.6%, respectively (13). According to the report of the National Lung Screening Trial (NLST), the mortality rate of LC reduced by 20% due to early screening (13). Therefore, early diagnosis not only provides a pathway for the adequate treatment to stop further progression of carcinogenic cells but also it is a potential solution to prevent LC casualties.

The existing methodologies of LC screening include X-ray and computed tomography (CT). However, the reliability of these techniques is a concern as the false positive rate was over 15% among these studies (14). Hence, in recent years, several alternative approaches (e.g., metabolomic, transcriptomic, genomic, and proteomic) for the identification of cancerous biomarkers have been explored for the early detection of LC so far. These approaches have been achieved through different pathological, molecular, and biochemical analyses, such as biopsy (supported by different methods: e.g., ctDNA, circulating tumor cells), sputum analysis, bronchoscopy, confocal micro-endoscopy, biomolecular markers (volatile, antibodies, etc.), endobronchial ultrasonography, optical coherence tomography, and positron emission tomography. This article demonstrates the potential and reliable pathway towards the early diagnosis of LC through these methodologies.

**Current approaches for LC diagnosis**

The two major diagnosis approaches to LC are (I) X-ray, and (II) computed tomography (CT) scan. Their methodologies, contributions, and limitations have been elucidated below.

**X-ray**

X-ray is one of the trivial methods for early LC detection. For LC detection, a chest X-ray is used, which includes an X-ray image of the heart, lungs, blood vessels, and bones (15). During the chest X-ray, a small amount of radiation beam passes through the body of the patient, and a black-and-white image is created on a film or computer (Figure 1A). Any unusual mass or nodules become visible on the X-ray image as the transmission of radiation varies due to the density of the cell. The main feature used in the X-ray image is the degree of contrast and the size of the nodules. However, in the early stage of LC, the nodules developed in the lungs are relatively small and show up as low-contrast white circular dots in the X-ray plates making the detection relatively hard (16). Again, the chest X-ray contains blood vessels and rib areas that overlap with the cancer mass or nodules, making them invisible in the X-ray image (17). Thus, X-ray is proved to be ineffective in early LC detection. Several recent studies have also investigated the effectiveness of X-ray in early LC detection. Rota et al. surveyed 765 subjects affected by LC (18). In their research, they found that the accuracy of X-ray based detection is ten times lower than other early-stage cancer detection techniques such as tomography. Gohagan et al. conducted another study to observe the effectiveness of X-ray in detecting different types of LC (19). The researchers found that out of 16 stage III–IV cancers, chest X-ray could only
detect nine cases. However, using deep learning methods, several research studies have been conducted to improve the accuracy of early LC detection using X-ray. Gordienko et al. presented that it is possible to remove the overlapping bone images from the X-ray image to improve the accuracy of detecting unusual mass and nodules in the chest X-ray using deep learning (20).

**CT scan**

Computed Tomography, also known as CT or CAT scan, is a specialized diagnostic method that uses a unique form of X-ray and computer technology to create medical images for soft tissues, bones, and internal organs (21). CT scan provides more detail information than traditional chest X-ray regarding early diagnosis of LC. CT scan produces cross-sectional images of the soft tissues in the chest and lungs area, which can be used to observe any unusual mass form or nodules (Figure 1B). Several researchers have demonstrated the efficiency of CT scans in detecting early-stage LC in their studies. Saghir *et al.* studied the CT scan of 4,104 smokers diagnosed with early LC and achieved an 83% detection rate (22). In another study, a total of 31,567 persons with the risk of LC have been diagnosed using a CT scan, and 484 participants have been diagnosed with cancer with a confidence rate of 95% (23). However, low-dose spiral CT scan images show pulmonary nodules, which is common among many people. This increases the chance of false-negative cases in the early detection of LC. Also, a high dose of radiation and high cost make CT scan less effective in early diagnosis (24). Recently, several new approaches have been accumulated with the traditional CT scan method to improve the accuracy of LC detection. Makaju *et al.* proposed a new approach to integrate image processing and machine learning technique to improve the accuracy of LC detection from CT scan images (25). In recent years, optical coherence tomography (OCT) is widely used, which is a special imaging technique. OCT uses low-coherence light to capture higher spatial resolution than CT and provides high-definition cross-sectional images of inner lungs, including the airway luminal wall. This can be used to detect any lesions in the bronchial, which can be considered as a sign of LC. Nonetheless, an alternative approach is needed to minimize the use of radiation and high cost in early-stage LC detection.

**Alternative approaches for LC diagnosis**

**Biopsy**

Lung biopsy is a test done to remove a tissue sample from the lung for examination. This small piece of the lung is taken with a special biopsy needle or during surgery. This type of biopsy is called a tissue biopsy, which is performed to detect different lung diseases, including LC. There are different types of biopsies available to detect lung diseases. For example, a needle biopsy (shown in Figure 2) is performed by putting a needle into the lung to take out a sample. Transbronchial biopsy is done with a bronchoscope, a long, thin tube with a tiny camera. Video-assisted thoracoscopic surgery (VATS) uses a special tiny camera called a thoracoscope, which transmits the chest image to the computer monitor. This is a minimally invasive process where the thoracoscope is put inside the chest cavity through a small incision. Contrarily, open biopsy involves a larger incision in the patient’s skin to reach and remove a small piece of the lung for further examination. These tissue biopsies have been widely used for LC diagnosis, but
the invasive nature limits their application, especially when repeated biopsies are needed. However, for early diagnosing LC, a minimally invasive procedure—liquid biopsy—aiming to primarily analyze circulating tumor cells (CTCs) and/or circulating tumor DNA (ct DNA) has gained interest from oncologists and basic researchers (shown in Figure 2). Nowadays, liquid biopsy is more feasible in clinical practice than before, because liquid biopsy is used to predict prognosis according to genetic alterations and monitor disease based on signature molecular markers in addition to the diagnosis of LC (27).

Traditionally, the histopathologic diagnosis of LC has been made based upon information obtained from tissue biopsies. A tissue biopsy can also help oncologists for the staging of LC; therefore, treatment modalities can be determined. However, tissue biopsy has the following limitations: (I) an invasive procedure, (II) an unreliable diagnostic technique in terms of tumor heterogeneity and metastasis at distant sites, and (III) lack of repeatability. These characteristics of tissue samples can demand further investigations on the patients. On the contrary, liquid biopsy has been demonstrated to be a viable surrogate for noninvasive assessment of tumor-specific biomarkers while diagnosing LC. Therefore, it can be potentially used for a variety of clinical and investigational applications (28). Several liquid biopsies (circulating biomarkers) can be investigated in blood, for example circulating cell-free tumor DNA (cf DNA), cell-free RNA (cf RNA), exosomes, tumor-educated platelets (TEP), and circulating tumor cells (CTCs) to make early detection of LC (28). However, identifying CTC and ct DNA for LC diagnoses are limited by the low concentrations found in blood at the early stages. However, the ability to study the genomic profile of cancer cells through the noninvasive sampling of blood or other body fluids represents one of the most exciting and rapidly improving fields in diagnosing LC (29). CTCs originate by cell detachment from the primary LC into circulation. Moreover, CTCs are considered to contribute to cancer progression and the development of metastases. CTCs have been isolated in blood of patients with LC at variable concentrations depending on cancer types and stages of diseases. Various studies have demonstrated a potential role of CTCs as prognostic biomarkers or in predicting and monitoring response to different treatments, including targeted therapies. Compared to ct DNA, CTCs allow comprehensive studies at the DNA, RNA, and protein levels as well as functional studies, including the establishment of cell lines and xenografts (29).
Circulating microRNAs for LC screening

MicroRNAs are a group of single-stranded, non-coding small RNA (length of ~22 nucleotides) (30,31), which affect the stability and rate of translation of their target messenger RNA (mRNA), thus regulate gene expression (30,32). There has been a significant alteration of patterns of miRNAs expressions during different developmental stages or in pathological conditions like cancer, cardiovascular diseases, etc. (33-35). Alteration of expression of miRNAs can be identified by Polymerase Chain Reaction (PCR), Northern Blotting method, microarray and deep sequencing method and can be applied for medical applications as well (31,36,37).

Abnormal levels of specific miRNAs can be identified in different stages, including early-stage, progression, or metastatic stages of cancer (31), and thus, these circulating small RNAs in plasma or serum can act as significant non-invasive biomarkers for diagnosis of LC in early stages (38). Wang et al. showed that differential expressions of miRNAs have been identified in various stages like diagnosis, treatment stage, and prognosis of NSCLC (Figure 3). The red color indicates increased expression of miRNAs in the peripheral blood of NSCLC patients, where blue color denotes the decreased expressions of miRNAs (31).

Yu et al. conducted a systematic review providing a summary of using circulating miRNAs as biomarkers for LC detection and indicated the diagnostic performance of miRNAs and miRNA panels in different cases (32). Yu et al. reviewed 17 studies in total, among which individual miRNA and miRNA panels were utilized as diagnostic biomarkers for both histological subtypes and stage-specific analysis. Considering the comparative analysis of sensitivity and specificity of both miRNA panels and individual miRNAs, the prior one appears better. However, no histology-specific miRNA could be identified because of limited changes in miRNA expressions in different histological subtypes. In the case of stage-specific analysis, both miRNA panels and individual miRNAs showed better efficacy for an advanced stage of LC diagnosis compared to early-stage with a minor difference in differential expressions of miRNAs in various stages. Boeri et al. conducted a prospective evaluation for early diagnosis of LC by CT screening trial and identified a panel of 15 miRNAs for predicting LC incidence in 2 years with sensitivity and specificity of 80% and 90%, respectively (39). Sozzi et al. conducted a similar study for prospective evaluation and validated a panel of 24 miRNAs, including the 15 miRNAs identified by Boeri et al. with similar sensitivity and specificity (40). Zaporozhchenko et al. [2016] identified a panel of miR-19b and miR-183 in plasma for histology variant LC detection with both sensitivity and specificity of 95% (41). Shen et al. also used a panel of miR-21, miR-486-5p, miR-126, and miR-210 for diagnosis of adenocarcinoma yielding 92% sensitivity and 97% specificity (42). While several miRNAs (e.g., miRNA-21, miRNA-155, etc.) are frequently reported in different studies, the higher frequency does not prove the better efficacy as a diagnostic marker because of their low median specificity. However, there is always a chance affecting the identification of miRNAs by disease conditions like COPD, asthma, tuberculosis, natural aging, gender, ethnicity, smoking behavior, etc. (43-47). Moreover, sample preparation, hemolysis of samples, and variation of
concentration of plasma and serum also affect the findings of miRNAs to a great extent (48-50). The most pragmatic approach is a combination of multiple individual miRNAs as a panel, which will enhance the diagnostic accuracy of circulating miRNAs for LC detection.

**Sputum analysis**

Sputum cytology is considered as the classic non-invasive screening method for assessing malignant and non-malignant diseases (51). Although it is quite a decent method with cost-effectiveness, it showed poor sensitivity and specificity for LC screening (52,53). In the early 1970s, NIH funded three randomized controlled trials that were conducted by the Mayo Clinic (54), Johns Hopkins University (55), and Memorial Sloan-Kettering (56) showing no benefit of sputum cytology as an effective outcome for the screening of LCs. Sputum cytology is not performed nowadays for early LC detection.

However, some recent studies showed that aberrant methylation of some genes persisting in the sputum of LC patients could be detectable by DNA methylation, avoiding the necessity of invasive screening procedures (57). Genetic alterations and epigenetic modifications are the earliest features of the pathogenesis of LC and DNA methylation, being one of the primeval epigenetic changes are associated with the manifestation and progress of LC (58). Liu et al. conducted a meta-analysis showing a mid-level accuracy of detection of methylated genes in sputum samples for early screening of LC with a sensitivity of 0.46 and specificity of 0.83 and they suggested to pick the superior genes as diagnostic biomarkers for LC (59). Since single DNA methylation has less efficacy as a biomarker for the detection of LC, some studies recommended combined methylated genes for improving the diagnostic value (60,61). Methylated FAM19A4 showed high sensitivity and methylated RASSF1A, FHIT, MGMT, and p16 showed a high positive likelihood ratio (PLR) suggesting that they have the potential of being a diagnostic marker for the screening test (62). Also, the methylation of FAM19A4, FHIT, and MGMT were reported to play the roles in the occurrence and deterioration of LC (63-66). Therefore, methylated SOX17, CDO1, ZFP42, TAC1, FAM19A4, FHIT, MGMT, p16, and RASSF1A are useful in the screening and auxiliary detection of LC.

**Detection of volatile organic compounds (VOCs) as biomarkers**

The differences between the levels of VOCs in different biofluids of a healthy person and cancer ailed patients can be a potential pathway to determine the existence and severity of LC. The major sources of these olfactory biomarkers are several biofluids, such as blood, plasma, interstitial fluid, exhaled breath, and urine. Horvath et al. suggest aroma(s) in the blood can be an appropriate screening approach for the detection of LC (67). Matsumura et al. demonstrated an ex-vivo analysis of urinary VOCs as biomarkers for LC (68). Liu et al. (shown in Figure 4A) found three key organic groups (ketones, alcohols, and benzene) in pleural samples where their derivatives were promising biomarkers.

**Figure 4** Volatile organic compounds (VOCs) as biomarkers for lung cancer diagnosis (A) GC/MS ion chromatograms of VOCs in benign (top) and cancerous (bottom) pleural samples, (B) 24 chemically reactive colorants used in colorimetric sensor for the detection of lung cancer from breath.
for cancer malignancy. Among 76 VOCs, nine of them including ethyl acetate, dichloromethane, n-heptane, xylene, ethyl-benzene, cyclo-hexanone, cyclo-hexanol, tetra-methyl benzene, and 2-ethyl-1-hexanol, are plausible compounds for screening to distinguish nonbenign from malignant pleural effusions. Due to apoptosis and necrosis, dead LC cells also origins of some specific VOCs, such as 1,3-di-tetrt-butylbenzene, 2,6-di-tetrtbutyl-1,4-benzoquinone, and n-decane (69).

Several analytical methods, such as GC-MS, GS-MS techniques with olfactometer (GS-MS-O), proton transfer-reaction MS (PTR-MS), selected ion flow tube MS (SIFT-MS), laser spectrometry and ion mobility spectrometry (IMS), electron impact mass spectroscopy (EIMS) integrated with two-dimensional gas chromatography (GC × GC) were employed for the detection of VOC biomarkers from urinary fluids, blood, plasma or interstitial fluids (70-74). However, e-noses are being widespread for their real-time and noninvasive detection from breath. Such form factors for chemi-resistive, optical, and electrochemical sensing approaches have been achieved for the miniaturization of e-noses (75-78). Obermeier demonstrated integrated e-noses with multiple amperometric sensors for the detection of VOC biomarkers, such as aldehyde, NO, and CO at sub-ppb levels for the diagnosis of LC (79). Mazzone’s group from Cleveland Clinic verified colorimetric based e-noses combined with a prediction model to diagnose LC, shown in Figure 4B (80). In their study, they collected 229 subjects’ data and diagnosed 92 individuals as carcinogenic patients with a diagnostic accuracy of 0.8 C-statistics. They also explored chemi-resistive based e-noses for the diagnosis of LC from respiration (81). They applied multivariate analysis methods for the sorting of benignity and malignancy of lung cells. Further, the support vector machine (SVM) was employed to generate an estimated model from the data they obtained. Their e-noses showed a sensitivity of 71.4% and 91.9% of specificity for the diagnosis of LC.

**Antibody-based biomarkers for early LC diagnosis**

Screening trials employing existing technologies, such as chest x-ray, CT, or sputum analysis, have far been incapable of reducing the mortality caused by LC. Most of these techniques suffer from inaccuracy and lower reliability due to the wrong diagnosis by false-positive errors caused by infection, benign tumors, pregnancy, and other factors (14). However, such doubtful deviations must need a reliable diagnosis method(s) that capable of distinguishing slow-growing preneoplastic lesions or benign lesions (14). Recently, the human serum has been broadly considered in the expedition for cancer biomarkers as blood is considered to deliver a dynamic representation of an individual’s pathophysiological condition.

Neuron-specific enolase (NSE) and lactate dehydrogenase (LDH) and have been acknowledged as prognosis markers for the SCLC (82). Contrarily, cytokeratin 19 fragment (CYFRA 21-1) has been identified as an independent biological factor for the NSCLC. Analyzing metadata of 2063 patients, CYFRA 21-1 was later corroborated as a possible biomarker for the diagnosis of NSCLC (83). Several antibody-based markers, such as carcinoembryonic antigen (CEA), cytokeratin 19 fragments (CYFRA21-1), carbohydrate antigen (CA) 125 and CA19, onecogene or tumor suppressor genes (e.g., Rb, p53, Bcl2), proliferation markers (Ki67), angiogenesis markers (vascular endothelial growth factor, VEGF), and different antigens (e.g., NSE, CA125, tissue polypeptide antigen, etc.) have been explored, but still needs sufficient proof to be recognized as establishing markers for the diagnosis of LC (84). Therefore, a very few prognosis markers have been examined to clinical trials as most of these biomolecules had low accuracy and specificity to diagnose LC in early-stage. However, recent research suggested that the antibodies against tumor-associated antigens (TAAs), called tumor-associated autoantibodies (TAABS), could be a possible pathway to early detection of LC (85). Compared with these traditional LC serological markers, TAABS offer exceptional advantages regarding the early diagnosis of LC, even in some cases 5 years ahead of regular diagnosis.

**Bronchoscopy**

Bronchoscopy is an invasive procedure to examine different conditions of the lungs by inserting a camera-connected lighted tube (Bronchoscope) into the lungs through a patient’s nose or mouth. As the inner tissues, bronchi, and bronchioles are exposed to the Bronchoscope, and it is possible to detect different anomalies in the lungs, including tumors, a sign of infection, excess mucus, etc. using bronchoscopy. There are two types of bronchoscopy—flexible bronchoscopy and rigid bronchoscopy. Rigid bronchoscopy employs a piece of larger and rigid equipment, which provides access to the proximal airways. It is generally performed for the removal of foreign substances and airway stents to control massive hemoptysis. It is also used for tumor debulking and dilation of the airways (86).
Contrarily, the flexible bronchoscopy uses a smaller and more flexible piece of equipment to access lower airways (e.g., third order of bronchi).

As bronchoscopy gives a detailed overview of different features of the lungs from inside, it is widely recommended for the diagnosis of early-stage cancer detection (87). In practice, fiber-optic bronchoscopy is widely used as it is safe (0.12% complication rate) and easy to perform (88). Tofolean et al. summarized the importance of bronchoscopy in early LC detection by comparing the performance of different bronchoscopy procedures with several traditional approaches such as biopsy, X-ray, spiral CT, etc. (89). In this study, researchers demonstrated that bronchial brush procedures in bronchoscopy could achieve positive results in 88–94% cases, which is higher than bronchial aspiration and has minimum complications than bronchial biopsy (90). De Roza et al. conducted a study among 226 patients to determine the success rate of bronchoscopy to detect early-stage LC (91). Among 226 patients, 84.5% of patients were successfully diagnosed with LC using bronchoscopy. However, the chance of false-negative is still high in flexible bronchoscopy, as small tumors inside the tissues cannot be detected in this procedure. Recent advancements and researches have addressed this shortcoming and proposed different techniques to improve the accuracy of cancer detection using bronchoscopy. For example, Herth et al. proposed narrow-band imaging (NBI) in flexible bronchoscopy procedures to improve the sensitivity and specificity of detecting early-stage LC (92). They performed the study among 62 patients and found that NBI can lower the sensitivity without impacting the accuracy of cancer detection using bronchoscopy. Among different bronchoscopy approaches, autofluorescence bronchoscopy and white light bronchoscopy are more prominent in practice.

Autofluorescence bronchoscopy (AFB) is a type of bronchoscopic procedure where flexible bronchoscopy is performed with a blue light instead of a white light (93). The main reason to use blue light is to differentiate the normal tissue and the malignant tissue by color without using any fluorescence-enhancing drug. AFB uses the same equipment used in regular flexible bronchoscopy except for a dedicated endoscopic system for capturing blue light imaging. As the captured images are of different wavelengths, in AFB, the normal tissues are visualized as green, and abnormal tissues are visualized as a reddish-brown color. For further inspection, normal bronchoscopic procedures are followed.

Compared to white light bronchoscopy (WLB), as shown in Figure 5, AFB offers low specificity in detecting early LC (94). Häussinger et al. performed a comparison study between AFB and WLB among 1,173 patients in Europe (95). Their research showed that sensitivity and specificity can be improved by combining WLB and AFB concerning WLB alone. In another study, Ueno et al. performed AFB among 31 patients to detect preinvasive bronchial lesions (early stage of invasive cancer) (96). Their results showed that AFB detected 64 lesions in the patients with a sensitivity of 94.7%, which is higher than the WLB (73.7%).

Figure 5 Different bronchoscopy procedures to identify LC in the same patient. (A) White-light bronchoscopy; (B) autofluorescence bronchoscopy image.
patients. In conclusion, using AFB with other existing methods, such as WLB or CT scans, can improve early-stage LC detection significantly.

**Endobronchial ultrasound (EBUS)**

EBUS is an effective procedure with a minimum invasion procedure to diagnose different lung diseases. EBUS uses ultrasound with regular bronchoscopy to visualize the inner tissues and airwall of the lungs (98). Due to low risk and high diagnostic value, EBUS is highly recommended in routine practice and early detection of lung diseases. There are two systems available for EBUS—linear EBUS and radial EBUS (88). For linear EBUS, the ultrasound transducer is used in the distal end, utilizing a fixed array to visualize the curvilinear pattern of the airways. In radial EBUS, a mechanical radial mini probe is used to visualize the properties of the peripheral of the lungs.

In recent years, researchers have studied the effectiveness of EBUS in detecting early-stage LC among patients. Herth *et al.* conducted a study among 648 patients to demonstrate the feasibility of EBUS in cancer detection (99). Their results showed that EBUS could improve the results of the regular bronchoscopic procedure without any complications in the patient's conditions. In a recent study, the same group researchers observed 105 different cases where patients have centrally located tumors in the lungs (100). EBUS methods detected all the cases with sensitivity and specificity of 89% and 100%, respectively. Miyazu *et al.* reported the effectiveness of EBUS in detecting early-stage LC in 16 different cases (101). Among these cases, EBUS has correctly diagnosed the depth of invasion. In another study, Takahashi *et al.* Used EBUS in detecting the degree of carcinoma invasion and found a 100% success rate in 14 cases (102). In conclusion, EBUS is proven as a highly effective procedure along with regular bronchoscopy to detect the depth and degree of invasion in the early stage of LC.

**Confocal endoscopy**

Confocal Endoscopy or confocal laser endoscopy (CLE) is an advanced imaging technique that permits real-time capturing of *in vivo* pictures of cellular and subcellular structures in the mucosa or living cells in lung tissue. CLE is a potential tool to explore alveolar elastic fiber structures, bronchus mucous membranes, and micro-vessels in the lung tissue. Hassan *et al.* studied 48 patients for the feasibility of CLE having with thinner probes to evaluate the malignancy of solitary pulmonary nodules. They employed r-EBUS coupled with CLE towards achieving 79.2% of accuracy (103). Fuchs *et al.* demonstrated a comprehensive study of 32 suspected patients who underwent CLE and bronchoscopy together. Their exploration showed significantly improved performance (sensitivity 96.0%, specificity 87.1%, accuracy 91.0%) to diagnose neoplastic changes (104). Su *et al.* verified needle-based CLE (nCLE) to compare with CT, EBUS, and X-ray, respectively (105). Their case studies exhibited that nCLE was viable to entree the extraluminal pulmonary nodules through a micro-needle for investigation with 91% accuracy. Comino *et al.* explored probe-based CLE (pCLE), integrated with computer-aided diagnoses (CAD) tools for the reliable identification of LC (106). Their structural graph analysis and computation methods were employed to deep-learning feature spaces to obtain enhanced CLE images (83.4% accuracy), which contain adequate visual information of neoplastic cell patterns of lung tissue.

**Future trends and conclusion**

Even with new technologies and treatments, LC is one of the major causes of patient mortality in the world. Several research studies showed that LC can be treated successfully if diagnosed at an early stage. One possible future solution to increase LC detection at an early stage is the use of artificial intelligence and machine learning. The available medical metadata of LC patients and treatments can be accurately visualize using different machine learning approaches (107). Furthermore, artificial intelligence can also be utilized to analyze patients' behavior, lifestyle, and food habits to detect possible LC patients at an early stage (107-109). Additionally, medical data from different patients can be analyzed using machine learning and artificial intelligence to propose a better treatment plan.

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**Footnote**

*Conflicts of Interest:* All authors have completed the ICMJE uniform disclosure form (available at http://dx.doi.org/10.21037/shc.2020.03.03). The authors have no conflicts of interest to declare.
**Ethical Statement:** The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

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